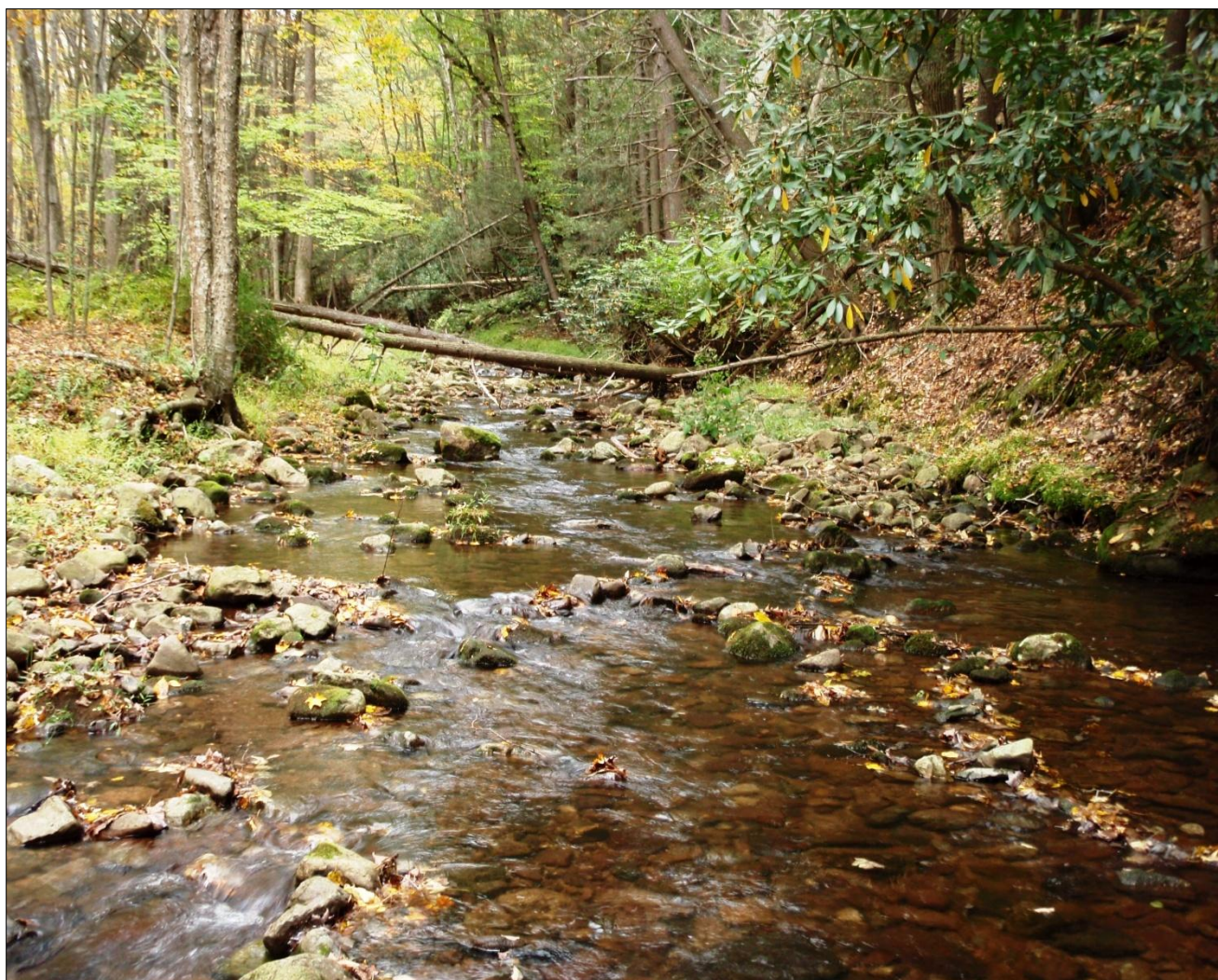




Integrity of benthic macroinvertebrate communities in Delaware Water Gap National Recreation Area

Eastern Rivers and Mountains Network 2008 summary report

Natural Resource Data Series NPS/ERMN/NRDS—2010/027



ON THE COVER

Vancampens Brook in Delaware Water Gap National Recreation Area.
Photograph by: Caleb Tzilkowski.

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All manuscripts in the series receive the appropriate level of peer review to ensure that the information is scientifically credible, technically accurate, appropriately written for the intended audience, and designed and published in a professional manner. Data in this report were collected and analyzed using methods based on established, peer-reviewed protocols and were analyzed and interpreted within the guidelines of the protocols.

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Acronyms

BCP	Boundary control point
BMI	Benthic macroinvertebrate
DEWA	Delaware Water Gap National Recreation Area
DO	Dissolved oxygen
ERMN	Eastern Rivers and Mountains Network
IDAS	Invertebrate Data Analysis System
MAHR	Mid-Atlantic Highlands Region
MBII	Macroinvertebrate Biotic Integrity Index
MTI	Macroinvertebrate Tolerance Index
NPS	National Park Service
RTH	Richest targeted habitat
SRMP	Scenic Rivers Monitoring Program
UNT	Unnamed tributary
USEPA	U.S. Environmental Protection Agency
USGS	United States Geological Survey

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Abstract

During 2008, the Eastern Rivers and Mountains Network (ERMN) of the National Park Service (NPS) began monitoring benthic macroinvertebrate (BMI) communities in wadeable streams throughout its nine parks. There were 23 randomly selected sites and 6 (non-random) targeted sites sampled throughout Delaware Water Gap National Recreation Area (DEWA) during October 2008. In addition to BMI samples, core water quality data (i.e., temperature, dissolved oxygen, pH, and conductivity) were collected and reach-scale habitat was characterized. Thirteen sites were located on streams where boundary control points (BCPs) have been established for water quality monitoring as part of the Scenic Rivers Monitoring Program (SRMP).

Core water quality parameters at DEWA sites were typical of forested watersheds with similar geologic characteristics. Relationships among core parameters were also typical – specific conductance generally decreased with decreasing pH, whereas dissolved oxygen concentrations (DO) consistently decreased with increasing water temperature. The pH of Little Flat Brook (8.66) was considerably higher than most DEWA streams, whereas pH was comparatively low (5.19) in UNT to Delaware River (Sunfish Pond). There were no exceptionally warm or cold streams throughout DEWA given the fall sampling period and DO concentrations were typically at or above saturation levels.

Benthic macroinvertebrate communities throughout DEWA streams had Macroinvertebrate Biotic Integrity Index values that ranged from 26.2 (UNT to Dingmans Creek) to 65.2 (Adams Creek). Taxa richness of BMI communities ranged from 19 (UNT to Delaware River/Sunfish Pond) to 37 (Little Bushkill Creek). Density of BMI was nearly three times as great in Little Flat Brook ($17,050 \text{ m}^{-2}$) as in the next highest stream (Sawkill Creek, $6,254 \text{ m}^{-2}$), whereas the lowest density of BMI was in Hornbecks Creek (319 m^{-2}).

Given that this report represented the first year of data collection, there were few inferences or management recommendations that could be confidently made. Biological communities (including BMI) can vary through time due to a range of naturally occurring biotic phenomena (e.g., interspecific competition, predation) and abiotic disturbances (e.g., severe drought, floods). It will take several years to determine the degree to which BMI communities naturally vary throughout DEWA and the remainder of the ERMN. Once natural variability of BMI communities is quantified, we will be in a better position to make inferences regarding the relative condition of sampled streams.

Introduction

During 2008, the Eastern Rivers and Mountains Network (ERMN) of the National Park Service (NPS) began monitoring benthic macroinvertebrate (BMI) communities in wadeable streams throughout its nine parks. This monitoring effort is a component of the ERMN Vital Signs monitoring program (Marshall and Piekielek 2007) as part of the nationwide NPS Inventory and Monitoring Program (Fancy et al. 2009).

One of the primary objectives of the ecological monitoring program in the ERMN is to evaluate status and trends in the condition of tributary watersheds flowing into and through member parks. Watershed condition is evaluated using measures of ecosystem integrity, including streamside bird species and communities (Mattsson and Marshall 2009), forest structure and composition (Perles et al. 2009), stream-dwelling benthic macroinvertebrates (Tzilkowski et al. 2009), stream chemistry, and watershed land use, type, and configuration (Marshall and Piekielek 2007). A primary purpose of the benthic macroinvertebrate monitoring protocol is to support the antidegradation or restoration of ERMN aquatic communities and their habitat (including water quality) by communicating monitoring program results to appropriate regulatory state and federal agencies.

Benthic macroinvertebrates are aquatic invertebrate animals larger than microscopic size that live on or within the stream bottom (benthos), and because they are a vital component of all functioning stream ecosystems, they are often used as indicators of ecosystem integrity. Types of BMI that are commonly used for water quality assessment include arthropods (insects, arachnids, and crustaceans), worms, clams, and snails. In addition to being instrumental to nutrient and carbon dynamics, BMI are an important link between basal resources (e.g., algae and detritus) and higher trophic levels (e.g., fish and birds) in stream food webs. Because BMI have been by far the most commonly used group for biological monitoring of aquatic ecosystems (Carter and Resh 2001), many metrics have been evaluated with respect to natural variation and responses to various sources of human-induced degradation. Given the proven ability to derive ecosystem integrity based on measures of BMI assemblage structure and composition, combined with the relatively low cost to sample, BMI are almost certainly the single best biological group to assess and monitor the ecological integrity of small and mid-sized streams.

At the time that this report was prepared, the BMI-monitoring protocol (Tzilkowski et al. 2009) had been developed, written, and received internal peer review, but had not undergone the final peer review process. This report was intended to provide preliminary results to natural resource managers at Delaware Water Gap National Recreation Area (DEWA) and at cooperating entities (e.g., Delaware River Basin Commission). The preliminary nature of data presented in this report should be considered prior to its use or dissemination.

Methods

Although a brief overview of the BMI monitoring methods is provided here, a detailed rationale of the sampling design and methods, in addition to Standard Operating Procedures, are provided in the BMI Monitoring Protocol (Tzilkowski et al. 2009). Much of this protocol is based on protocols developed by the U.S. Geological Survey ([USGS] Moulton et al. 2000, Moulton et al. 2002) and Bowles et al. (2006) because those protocols and programs have already undergone considerable evaluation and revision. We modified those protocols to fit the character of ERMN parks and anticipated monitoring resources.

Site Selection

There are two types of sampling sites in the BMI Monitoring Program – probabilistic (i.e., stratified-random) sites and non-random “targeted” sites. The probability-based design was developed by Mattsson and Marshall (2009) for the ERMN Streamside Bird Monitoring Program. This design was adopted for the BMI Monitoring Program for several reasons: (1) the design provided a population of wadeable stream sizes (i.e., generally 2nd to 4th Strahler stream order) that were suited to sampling methods and metrics that have been thoroughly developed and tested; (2) the population of “medium-sized” streams were more hydrologically stable than smaller intermittent streams and more safely and consistently accessible than larger rivers; and (3) collocation of BMI sites with streamside bird and water quality monitoring sites will provide multiple lines of evidence, with both terrestrial and aquatic components, to better evaluate trends in ecosystem condition at a landscape scale.

There were 23 randomly selected sites and six targeted sites throughout DEWA (Table 1; Figure 1) in 2008. Four stream sites did not have surface flow during the 2008 sampling season but will be revisited during 2009 to determine if they are typically dry during the fall. Those streams were Conashaugh Creek, Dry Brook, unnamed tributary (UNT) to Adams Creek, and UNT to Delaware River (Namanock Island). If these streams are consistently dry during the fall, other sites from the probability-based sample population may be substituted in their place. There are 13 streams throughout DEWA where boundary control points (BCPs) have been established for water quality monitoring as part of the Scenic Rivers Monitoring Program (SRMP). Water quality in these streams has been, and will likely continue to be, routinely monitored by DEWA staff to establish special protection regulations for water quality in these tributaries. Seven of the 13 streams with BCPs were coincidentally selected as probabilistic sites. The remaining six streams were chosen as targeted sites. Although the sites were not always directly collocated with BCPs, the BMI data will likely be used to support the SRMP water quality monitoring effort. An additional six targeted sites were chosen on streams where BCPs were established, but not selected using the probabilistic strategy – these sites were chosen because they will support the SRMP.

Field Methods

The sampling unit for the BMI monitoring program is the stream reach, which for the ERMN program is defined as a length of stream chosen to represent a uniform set of physical, chemical, and biological conditions within a stream segment. The length of sampled reaches differs among watersheds but their length is proportional (i.e., 40 ×) to stream width. Tributary reaches within floodplains of large rivers (e.g. Delaware River) were typically not considered as candidate sites because those areas were thought to exhibit considerable natural variation.

Table 1. Types of benthic macroinvertebrate monitoring sampling sites throughout Delaware Water Gap National Recreation Area. Probabilistic sites were chosen with a stratified-random design and collocated with Streamside Bird monitoring sites whereas boundary control points (BCP) were previously defined Scenic Rivers Monitoring Program water quality monitoring sites. Sampling locations at BCP sites that were not chosen with the probabilistic method were considered “Targeted” sites.

Stream	Probabilistic sites	BCP	Targeted
Adams Creek	X	X	
Big Flat Brook		X	X
Mill Creek	X		
Bushkill Creek		X	X
Caledonia Creek	X		
Conashaugh Creek	X		
Dingmans Creek	X	X	
Dry Brook	X		
Dunnfield Creek	X		
Fuller Brook	X		
Hornbecks Creek	X	X	
UNT* to Adams Creek	X		
Little Bushkill Creek	X	X	
UNT* to Dingmans Creek	X		
Little Flat Brook		X	X
UNT* to Toms Creek	X		
Raymondskill Creek	X	X	
UNT* to Delaware River (Namanock Island)	X		
Sawkill Creek		X	X
Shimers Brook		X	X
Slateford Creek	X		
Spackmans Creek	X		
UNT* to Delaware River (Arrow Island)	X		
UNT* to Delaware River (Sunfish Pond)	X		
Toms Creek	X	X	
Vancampens Brook	X	X	
Van Campen Creek	X		
Vandermark Creek		X	X
White Brook	X		

*UNT = Unnamed tributary

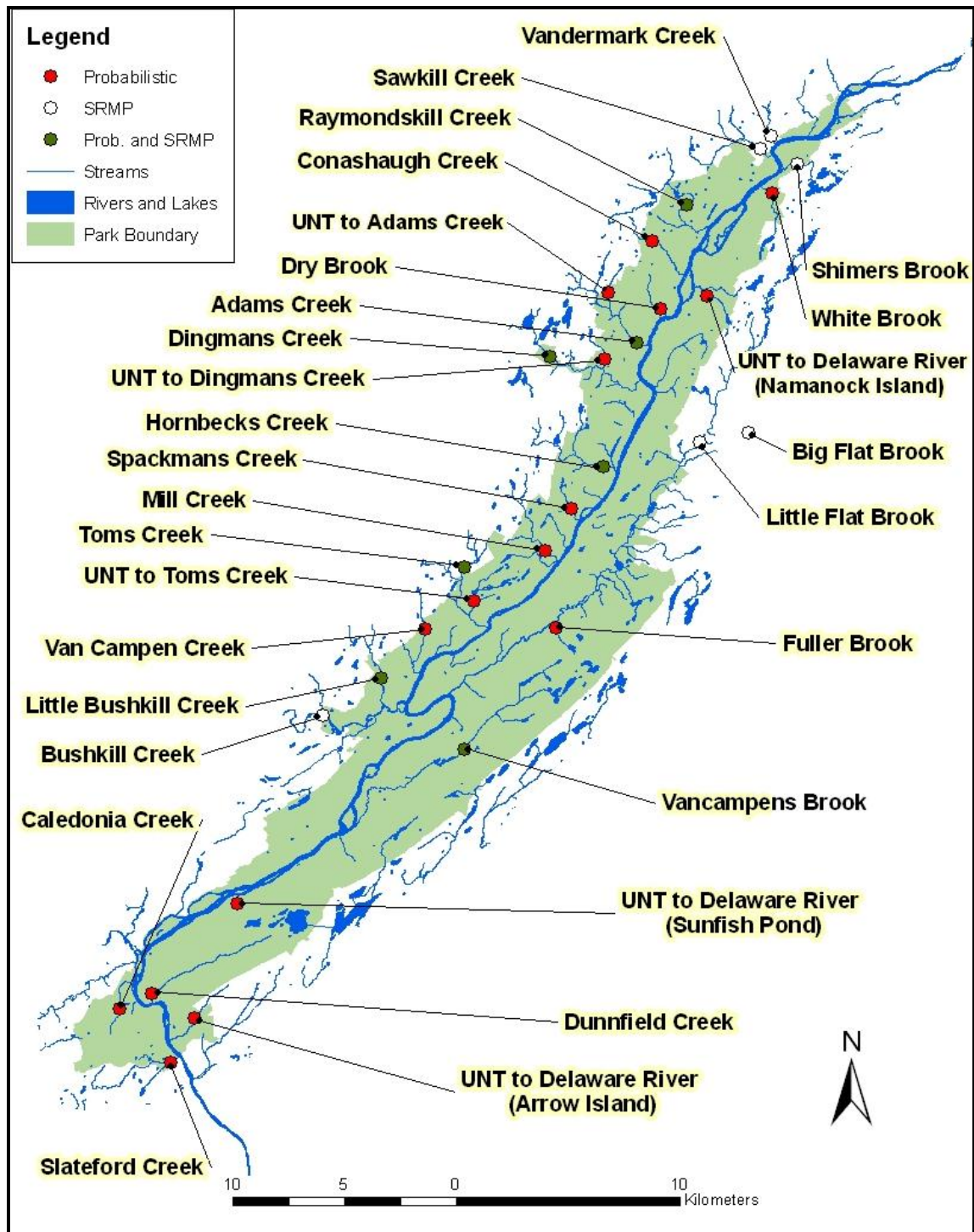


Figure 1. Benthic macroinvertebrate sampling sites throughout Delaware Water Gap National Recreation Area. Probabilistic sites (red circles) were chosen with a stratified-random design and collocated with Streamside Bird monitoring sites. Targeted sites (white circles) were selected to augment DEWA water quality monitoring, which is part of the Scenic Rivers Monitoring Program (SRMP). Some SRMP sites were coincidentally included in the probability-based sampling design (green circles).

Because the probabilistic site selection strategy was developed for the Streamside Bird Monitoring Program, these BMI sites were collocated with Streamside Bird monitoring sites and anchored on the point count location that is closest to the point of access (either upstream-most or downstream-most point count). Sampling is conducted within Streamside Bird transects; therefore, sampling progresses either upstream or downstream into transects, depending on whether the anchor point is at the upstream- or downstream-most end of the transect.

Sampling was conducted during October 2008. The ERMN method for collecting BMI throughout DEWA is termed semi-quantitative richest-targeted habitat (RTH, Moulton et al. 2002) sampling which is a type of disturbance-removal sampling. Although similar to more common kick sampling methods, RTH sampling calls for consistent and thorough collection of BMI from a fixed area; thus, it is considered a more precise method and allows for estimation of stream productivity unlike many other sampling methods. Many BMI disturbance sampling methods are qualitative (not quantitative) and are comparatively inconsistent because there is no measurement of sampling area – instead, those methods usually rely on a timed sampling effort. For the RTH method, five discrete samples are collected from riffles throughout the reach and are ultimately composited into a single homogenous sample. Ideally, discrete samples are taken from different riffles, but if fewer than five riffles are present, samples may be taken from the same riffle. Physical conditions (i.e., depth, flow, and substrate) are recorded at each sampling location and are as similar as possible among replicates. Sampling is conducted by defining a 0.25 m² sampling area with a template and then disturbing substrate within that area so that BMI are dislodged and then drift into a net placed downstream of the sampling area. The composited samples yield 1.25 m² of sampled area at each site.

In addition to BMI samples, core water quality data (i.e., temperature, dissolved oxygen, pH, and specific conductance) were collected and reach-scale habitat is characterized using the U.S. Environmental Protection Agency rapid bioassessment method (Barbour et al. 1999). Samples are processed in the field by using an elutriation method to remove mineral materials and large organic matter (e.g., whole leaves and sticks). Samples are preserved in 95% ethanol, packed carefully, and transported to the laboratory for processing and identification.

Laboratory Methods

Laboratory methods for processing samples in the ERMN BMI Program rely a great deal on procedures developed by the USGS (Moulton et al. 2000). A fixed-count subsample of $300 \pm 20\%$ individuals are sorted and identified from each sample. The relatively large subsample size yields data that meets quality standards (i.e. precision and accuracy) required by most monitoring programs; however, processing and identifying additional individuals (> 300) does not typically yield enough additional information to justify the added effort (Moulton et al. 2000). Generally, BMI were identified to genus using standard dichotomous keys, but some groups (e.g., Chironomidae, Oligochaeta) were identified to coarser taxonomic levels. Microsoft Access 2007 is the primary software used for storing and managing ERMN BMI and stream habitat data, whereas the Invertebrate Data Analysis System (IDAS *version 5*, U.S. Geological Survey, Raleigh, NC) was used for resolving taxonomic ambiguity issues and calculating metrics that describe the structure and diversity of BMI communities.

Data Analysis

We calculated all BMI community metrics with IDAS and calculated the Macroinvertebrate Biotic Integrity Index (MBII; Klemm et al. 2003) using Microsoft Excel 2007. The MBII was developed by the U.S. Environmental Protection Agency's (USEPA) Environmental Monitoring and Assessment Program and was ultimately used for the USEPA's Wadeable Stream Assessment (USEPA 2006, Herlihy et al. 2008).

The rationale behind biotic integrity indices is that a suite of metrics that represent community structure, pollution tolerance, functional feeding groups and habitat occurrences, life history strategies, disease, and density provide insights regarding how biological communities respond to different natural and anthropogenic stressors (Klemm et al. 2003). A common stream bioassessment practice is to compare BMI community metrics from candidate streams to the same metrics from reference streams. Reference streams are "least disturbed," similarly sized streams within comparable geographic and geologic settings that provide an estimate of least-impaired stream communities. Departure of the sampled BMI community from expected BMI community composition (i.e., reference streams) serves as a measure of stream impairment. The MBII is one such index that uses reference streams to assess stream impairment.

The MBII was chosen for use in the ERMN because it was developed for upland and lowland streams dominated by riffle habitat in the Mid-Atlantic Highlands Region (MAHR). Moreover, the MBII was based on a large dataset of 574 wadeable stream reaches and was thoroughly tested. The MBII is a broadly applicable measure of stream impairment because it is based on several factors that affect aquatic communities throughout the MAHR. Impaired and reference streams for the MBII were identified by Klemm et al. (2003) using water chemistry, qualitative habitat, and minimum organism count criteria. Impaired reaches were defined by meeting any one of the following criteria: pH <5, chloride >1000 µeq/L, sulfate >1000 µg/L, total phosphorous >100 µg/L, total nitrogen >5000 µg/L, or a mean qualitative habitat score <10 (of a possible 20). Reference reaches met all of the following criteria (Klemm et al. 2003): sulfate <400 µg/L, Acid Neutralizing Capacity (ANC) >50 µeq/L, chloride <100 µeq/L, total phosphorous <20 µg/L, total nitrogen <750 µg/L, mean qualitative habitat score >15, and at least 150 organisms.

The MBII uses seven metrics selected from the 100 that are commonly used by governmental agencies throughout the MAHR. The metrics chosen were those that performed best in terms of range, precision, responsiveness to various human-induced disturbances, relationship to catchment area, and redundancy (Table 2; Klemm et al. 2003). Most MBII metrics are counts or proportions of taxa in the community that are characterized as tolerant or intolerant to human perturbations; however, one of the metrics (Macroinvertebrate Tolerance Index; MTI) is more complex because it incorporates values (0–10) for each taxon with respect to pollution tolerance, weighted by taxon abundance, and results in higher scores as the proportion of taxa tolerant to general pollution increases (Klemm et al. 2003). Pollution Tolerance Values (PTV) incorporated in the MTI were average tolerances to "various types of stressors" (Klemm et al. 2002).

Table 2. Macroinvertebrate Biotic Integrity Index metric descriptions and their directions of response to increasing human perturbation (Response) from Klemm et al. (2003).

Metric	Description	Response
Ephemeroptera richness	Number of Ephemeroptera (mayfly) taxa	Decrease
Plecoptera richness	Number of Plecoptera (stonefly) taxa	Decrease
Trichoptera richness	Number of Trichoptera (caddisfly) taxa	Decrease
Collector-filterer richness	Number of taxa with a collecting or filtering-feeding strategy	Decrease
Percent non-insect individuals	Percent of individuals that are not insects	Increase
Macroinvertebrate Tolerance Index	$\sum_i p_i t_i$, where p_i is the proportion of individuals in taxon i and t_i is the pollution tolerance value (PTV) for general pollution	Increase
Percent five dominant taxa	Percentage of individuals in the five numerically dominant taxa	Increase

We also present three other commonly used BMI community metrics (taxa richness, Shannon's Diversity and Evenness) for comparison because they are likely to be familiar to most readers of this report. Taxa richness was the combined number of unique taxa (usually genera). Shannon's diversity and evenness were calculated with IDAS using formulae provided by Brower and Zar (1984), which were:

Shannon's Diversity (H'): information theory-based index that measures the "uncertainty" of a taxon selected at random from the community. High diversity is associated with high uncertainty and low diversity with low uncertainty. This index is the equivalent of the Brillouin's diversity index, but it is intended for use when the abundance data come from a random sample of the community or subcommunity.

$$H' = (N \log_{10} N - \sum n \log_{10} n) / N$$

Shannon's Evenness (J'): ratio of the observed Shannon diversity to the maximum possible diversity (that is, diversity when individuals are distributed as evenly as possible among the species). Like the Shannon diversity index, this measure is intended to be used when the abundance data come from a random sample or the community or subcommunity

$$J' = H' / H_{\max}' \text{ where } H_{\max}' = \log_{10} S$$

Abbreviations used in formulae: S = number of taxa in sample, n = abundance of an individual taxon, N = total number of individuals in sample.

Results

Benthic Macroinvertebrate Communities

Benthic macroinvertebrate communities throughout DEWA streams had MBII values that ranged from 26.2 (UNT to Dingmans Creek) to 65.2 (Adams Creek, Figure 2). Streams that were part of the SRMP were distributed throughout the gradient of streams. Total taxa richness ranged from 19 (UNT to Delaware River/Sunfish Pond) to 37 (Little Bushkill Creek, Table 3). Streams with the top four MBII scores, as expected, had the highest combined Ephemeroptera, Plecoptera, and Trichoptera (EPT) scores which ranged from 20 to 23. Only two other streams (Sawkill Creek and Spackmans Creek) had EPT scores as high as 20 – most EPT scores were between 12 and 17. The lowest EPT score was from the UNT to Dingmans Creek, which also had the lowest overall MBII score. The collector-filterer taxa richness metric was relatively high (8) for the top two MBII-ranking streams but there was not a consistent pattern (e.g., decline) among the other streams. The proportional metrics (%Non-insects and %5 dominant) and Shannon diversity and evenness metrics generally responded as expected – with increasing MBII scores, the proportional and Shannon metrics decreased and increased, respectively. Finally, as expected, the MTI decreased with increasing MBII scores and ranged from 3.27 to 4.91.

Density of BMI was nearly three times as great in Little Flat Brook ($17,050 \text{ m}^{-2}$) as in the next highest stream (Sawkill Creek, $6,254 \text{ m}^{-2}$; Figure 3). Other density differences among streams were comparatively modest, but there appeared to be a general pattern among streams. There were four pairings of streams that had similar BMI densities. Those pairs were streams with BMI densities of: 1) $\approx 6,000 \text{ m}^{-2}$ (Sawkill Creek and Raymondskill Creek); 2) $\approx 4,000 \text{ m}^{-2}$ (Bushkill Creek and Big Flat Brook); and 3) $\approx 2,500 \text{ m}^{-2}$ (Mill Creek and Little Bushkill Creek). Densities of BMI in the remaining streams were comparatively low and ranged from 319 to $1,877 \text{ m}^{-2}$.

Water Quality

Physical and chemical characteristics can vary markedly, both daily and annually. Although there are limitations to point-in-time characterizations of core water quality parameters, these measures can be helpful when evaluating patterns in biological data; moreover, extreme changes to these parameters can sometimes be detected with point-in-time samples. Core water quality parameters (pH, specific conductance, temperature, DO) at DEWA sites were typical of forested watersheds with similar geologic characteristics. Relationships among core parameters were also typical – specific conductance generally decreased with decreasing pH (Figure 4), whereas DO concentrations consistently decreased with increasing water temperature (Figure 5). The pH of Little Flat Brook (8.66) was considerably higher than most DEWA streams, whereas pH was comparatively low (5.19) in UNT to Delaware River (Sunfish Pond). Other than those two streams, pH ranged between 6.15 and 8.24. One interesting observation was that specific conductance at the UNT to Dingmans Creek was high (226 mg/L) compared to streams with similar pH values. There were no exceptionally warm or cold streams throughout DEWA given the fall sampling period and DO concentrations were typically at or above saturation levels.

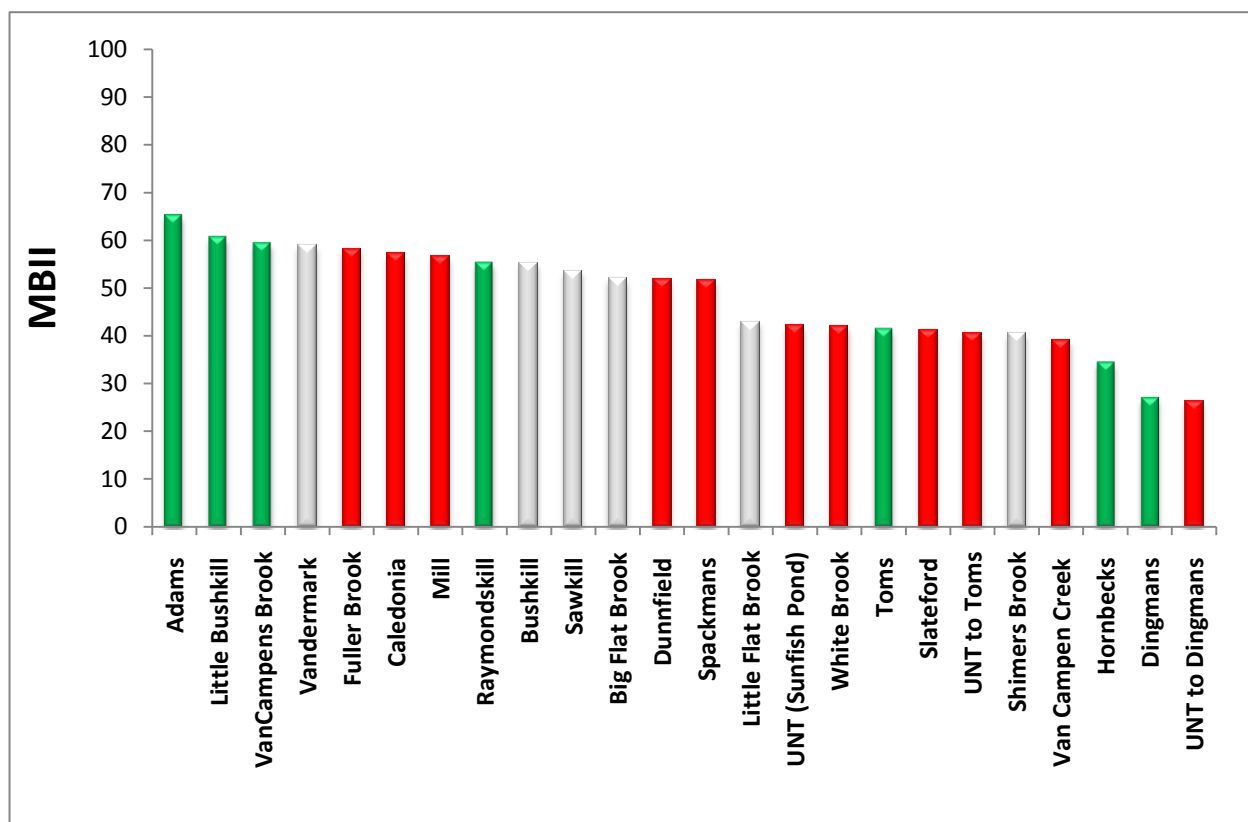


Figure 2. Macroinvertebrate Biotic Integrity Index (MBII, Klemm et al. 2003) values for benthic macroinvertebrate samples collected at sampling sites throughout Delaware Water Gap National Recreation Area in October 2008. Probabilistically chosen sites located on streams that were also part of the Scenic Rivers Monitoring Program (SRMP) are green bars whereas sites that were part of SRMP, but not probabilistically chosen, are white bars. The remaining sites (red bars) were probabilistically chosen but not part of the SRMP.

Table 3. Summary metrics and multimetric indices for benthic macroinvertebrate communities sampled from Delaware Water Gap National Recreation Area during October 2008. Direction of metric or index response to increasing stream ecosystem integrity are denoted parenthetically by + or -. Richness metrics included total taxa richness (Total), and richness of Ephemeroptera (E), Plecoptera (P), Trichoptera (T), and Collector or Filter feeders (C-F). Proportional metrics included the percent of individuals in samples that were non-insect taxa (%Non-insects) or that comprised the combined five dominant taxa in the community (%5 dominant). Indices were the Macroinvertebrate Tolerance Index (MTI) and the Macroinvertebrate Biotic Integrity Index (MBII). Probabilistically chosen sites located on streams that were also part of the Scenic Rivers Monitoring Program (SRMP) are denoted by **. Streams that were part of SRMP, but not probabilistically chosen, are denoted by *.

Stream	Richness (+)					Proportional (-)		Shannon (+)		Indices	
	Total	E	P	T	C-F	%Non-insects	%5 dominant	Diversity	Evenness	MTI (-)	MBII (+)
Adams Creek**	35	7	5	11	8	1	63	1.19	0.77	3.50	65.2
Little Bushkill Creek**	37	8	2	11	8	10	50	1.28	0.82	3.68	60.6
Vancampens Brook**	33	6	5	11	4	0	53	1.26	0.83	3.48	59.4
Vandermark Creek*	30	5	6	9	5	5	50	1.27	0.86	3.71	59.0
Fuller Brook	29	4	6	8	3	0	55	1.20	0.82	3.57	58.2
Caledonia Creek	34	6	6	7	4	2	61	1.21	0.79	3.27	57.1
Mill Creek	28	7	3	7	5	2	56	1.22	0.84	3.65	56.6
Raymondskill Creek**	34	8	3	7	7	11	42	1.32	0.86	4.34	55.3
Bushkill Creek*	30	8	2	8	5	5	57	1.18	0.80	3.73	55.2
Sawkill Creek*	31	7	5	8	4	0	73	1.00	0.67	3.32	53.5
Big Flat Brook*	33	6	2	8	7	6	67	1.11	0.73	3.78	51.9
Dunnfield Creek	33	5	5	8	3	4	57	1.21	0.80	3.83	51.8
Spackmans Creek	33	8	4	8	4	10	49	1.27	0.84	3.92	51.5
Little Flat Brook*	24	5	2	7	4	5	73	1.02	0.74	3.60	42.9
UNT to Delaware River	19	1	5	5	2	5	68	1.06	0.83	3.63	42.2
White Brook	20	2	3	5	3	1	68	1.04	0.80	3.46	41.8
Toms Creek**	29	5	5	8	5	20	54	1.22	0.84	4.14	41.5
Slateford Creek	28	3	4	6	4	9	63	1.12	0.78	3.40	41.1
UNT to Toms Creek	28	7	3	8	5	6	72	1.00	0.69	4.52	40.5
Shimers Brook*	26	2	2	8	6	1	83	0.89	0.63	4.38	40.4
VanCampen Creek	30	5	3	9	5	13	61	1.11	0.75	4.17	39.0
Hornbecks Creek**	29	5	4	7	5	32	68	1.07	0.73	4.07	34.5
Dingmans Creek**	24	4	2	7	8	57	81	0.91	0.66	4.91	27.1
UNT to Dingmans Creek	24	1	2	6	6	34	65	1.10	0.79	4.91	26.2

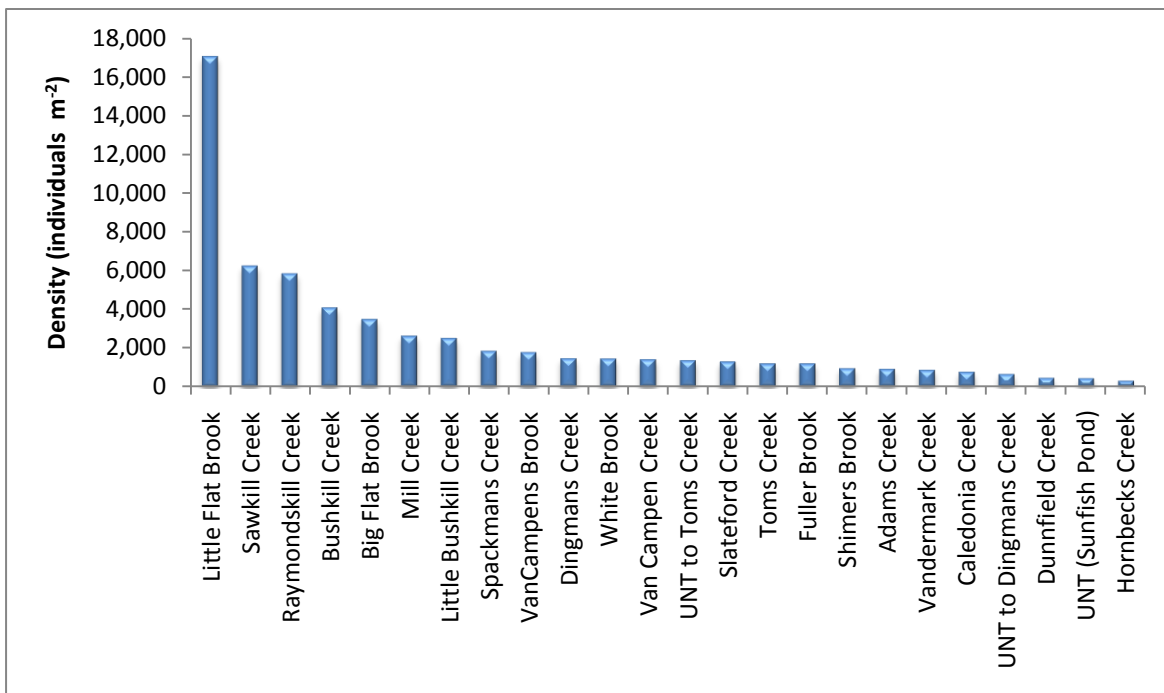


Figure 3. Density (individuals/m²) of benthic macroinvertebrates collected at sampling sites throughout Delaware Water Gap National Recreation Area (DEWA) in October 2008.

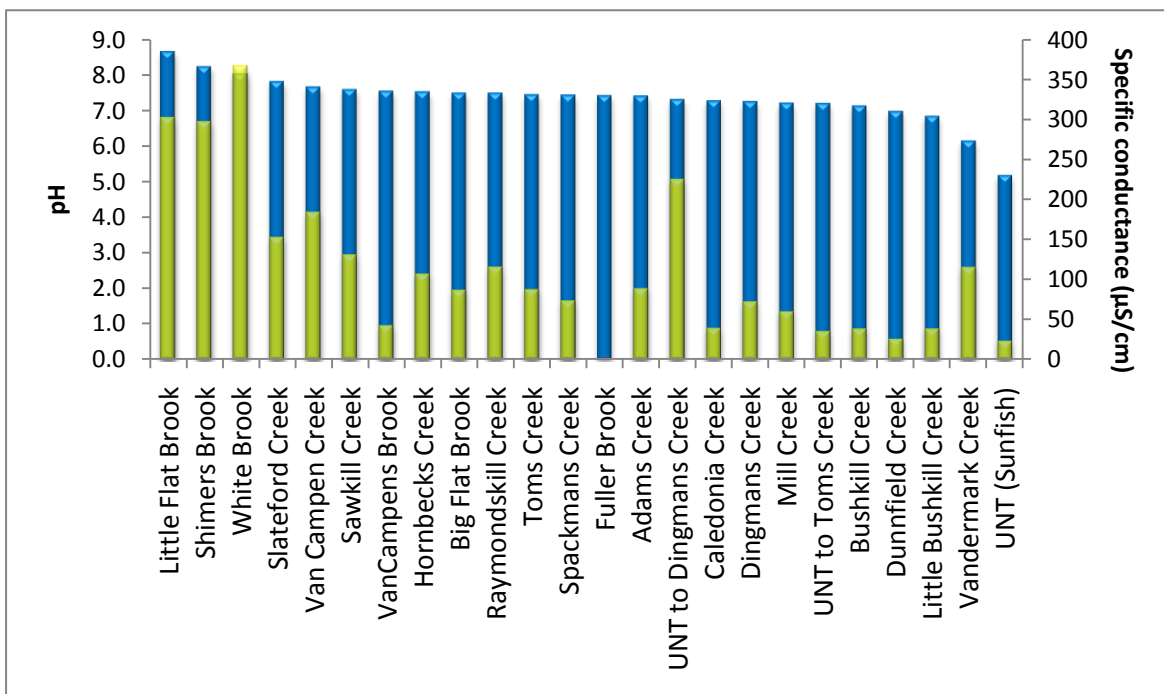


Figure 4. pH (blue bars) and specific conductance (yellow bars) of water at sampling sites throughout Delaware Water Gap National Recreation Area (DEWA) in October 2008. Specific conductance data were missing for the unnamed tributary (UNT) to Flat Brook.

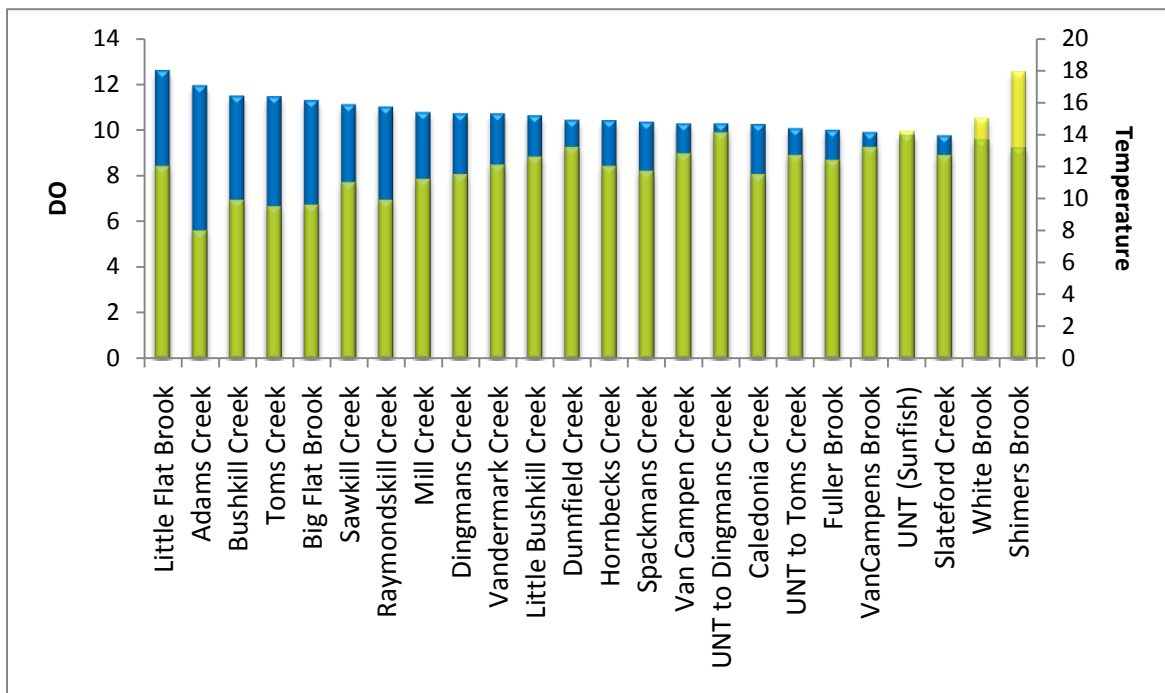


Figure 5. Dissolved oxygen concentration (blue bars) and temperature (yellow bars) of water at sampling sites throughout Delaware Water Gap National Recreation Area in October 2008.

Discussion

This report summarized results from the first sampling season of the ERMN BMI monitoring program at DEWA. The effort was largely successful in that it provided quality data for the majority of selected sites – four selected sites were not sampled due to lack of surface flow. All components of the protocol worked well. This was not a surprise because they were based largely on widely used USGS protocols. The primary challenge to interpreting the data (as discussed in the methods section) was that, because the ERMN protocol did not precisely follow other state or regional protocols, comparing our data with other efforts included qualifications.

Given that this report represented the first year of data collection, there were few inferences or management recommendations that could be confidently made. Biological communities (including BMI) can vary through time due to a range of naturally occurring biotic phenomena (e.g., interspecific competition, predation) and abiotic disturbances (e.g., severe drought, floods). It will take several years to determine the degree to which BMI communities naturally vary throughout DEWA and the remainder of the ERMN. Once natural variability of BMI communities is quantified, we will be in a better position to make inferences regarding the relative condition of sampled streams.

With each future sampling season, the ERMN BMI monitoring program will be refined and improved. It is anticipated that metrics and indices will be calibrated so that more precise and accurate comparisons can be made between DEWA streams and streams throughout the region. There were obvious natural differences among DEWA stream types, most obviously whether they could be broadly defined as “limestone” or “freestone” streams. Along with a variety of other differences, limestone streams generally have much higher pH and conductivity than freestone streams. These differences result in different biological communities including BMI; consequently, metrics and indices can be calibrated for different stream types. The abundance of BMI in Little Flat Brook demonstrated how limestone streams can be (presumably) naturally different than freestone streams. In the future, metrics and indices will be catered to DEWA stream “types,” which the ERMN will use in combination with spatial data to make better assessments regarding stream ecosystem condition at a park scale.

In addition to calibrating the MBII and its constituent metrics, the ERMN will add other measures of stream integrity as more data are gathered. For example, another meaningful way to express BMI community condition is with Observed/Expected Indices that estimate the number of taxa (e.g., genera) that have been lost (i.e., extirpated) from a given stream (Yuan 2008). To use these methods, the expected number of taxa for a given stream type must be established from the least-disturbed streams in the region (i.e., DEWA). This process will likely begin after next season when assessments regarding natural variability of BMI communities can be at least coarsely made.

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